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Supporting document

Risk and technical assessment – Application A1228

Endo-1,4-beta-xylanase from GM *Trichoderma reesei* (gene donor: *Talaromyces leycettanus*) as a processing aid

Executive summary

Novozymes Australia Pty Limited applied to Food Standards Australia New Zealand (FSANZ) to amend Schedule 18 of the Australia New Zealand Food Standards Code (the Code) to include endo-1,4-beta-xylanase (EC 3.2.1.8) as a processing aid for use in brewing, fats and oils processing, grain processing and potable alcohol production. The enzyme is sourced from a genetically modified (GM) strain of *Trichoderma reesei (T. reesei)* containing the endo-1,4-beta-xylanase gene from *Talaromyces leycettanus*.

FSANZ has undertaken an assessment to determine whether the enzyme achieves its technological purpose in the quantity and form proposed to be used and to evaluate any public health and safety concerns that may arise from its use.

FSANZ concludes that the proposed use of the enzyme in brewing, fats and oils processing, grain processing and potable alcohol production is technologically justified for use at levels consistent with Good Manufacturing Practice. Analysis of the evidence provides adequate assurance that the use of this enzyme, in the quantity and form proposed, is justified.

No public health and safety concerns were identified in the assessment of endo-1,4-betaxylanase produced by this GM *T. reesei* under the proposed use conditions. *T. reesei* has a long history of safe use as a production microorganism of enzyme processing aids, including several that are already permitted in the Code. The production organism is neither pathogenic nor toxigenic and a biotechnology assessment confirmed the presence and stability of the inserted DNA. Bioinformatics analysis indicated that the produced endo-1,4beta-xylanase does not demonstrate substantial homology with any known toxins or food allergens.

Toxicity testing of the endo-1,4-beta-xylanase enzyme showed no evidence of genotoxicity *in vitro* and the no observed adverse effect level (NOAEL) in a 90-day oral gavage study in rats was the highest dose tested, 1051 mg total organic solids (TOS)/kg body weight (bw)/day. The theoretical maximum daily intake (TMDI) was calculated as 0.15 mg TOS/kg body weight/day. Comparison of the NOAEL and the TMDI results in a Margin of Exposure (MOE) of around 7000.

Based on the reviewed data it is concluded that in the absence of any identifiable hazard an Acceptable Daily Intake (ADI) 'not specified' is appropriate.

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1 Introduction

Novozymes Australia Pty Limited (Novozymes) applied to Food Standards Australia New Zealand (FSANZ) to permit the use of the enzyme, an endo-1,4- β -xylanase (EC 3.2.1.8), as a processing aid for use in brewing, fats and oils processing, grain processing and potable alcohol production. The enzyme will be referred to in this report as endo-1,4-beta-xylanase.

This enzyme is sourced from a genetically modified (GM) strain of *Trichoderma reesei* (*T. reesei*) containing the endo-1,4-beta-xylanase gene from *Talaromyces leycettanus* (*T. leycettanus*). The host organism was developed from *T. reesei* RUT-C30 and derived from the original isolate from QM6A (ATCC 13631, Seidl et al. 2008).

1.1 Objectives of the assessment

Schedule 18 of the Australia New Zealand Food Standards Code (the Code) contains permission for enzymes of microbial origin including endo-1,4- β -xylanase (EC 3.2.1.8) from *T. reesei*. Currently, there is no permission in section S18-9 for endo-1,4-beta-xylanase produced by a GM strain of *T. reesei* containing the gene for endo-1,4-beta-xylanase from *T. leycettanus*. Assessment is required before permission can be considered for its use as a processing aid.

The objectives of this risk and technical assessment were to:

- determine whether the enzyme provides the proposed technological purpose as a processing aid in the quantity and form proposed to be used
- evaluate potential public health and safety concerns that may arise from the use of this enzyme as a processing aid, specifically by considering the
 - history of use of the gene donor and production microorganisms
 - characterisation of the genetic modification(s)
 - safety of the enzyme.

2 Food technology assessment

2.1 Characterisation of the enzyme

The production microorganism of the endo-1,4- β -xylanase enzyme is a GM strain of *T. reesei.* The applicant provided information regarding the identity of the endo-1,4- β -xylanase enzyme. FSANZ verified this by using the IUBMB¹ enzyme nomenclature database (McDonald et al. 2009). Details of the identity of the enzyme are provided in Table 1.

¹ International Union of Biochemistry and Molecular Biology

2.1.1 Identity of the enzyme

Table	1	Identity
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Generic common name:	Xylanase
Accepted IUBMB name:	Endo-1,4-β-xylanase
Systematic name:	4-β-D-xylan xylanohydrolase
Other names:	endo- $(1\rightarrow 4)$ - β -xylan 4-xylanohydrolase; endo-1,4-xylanase; xylanase; β -1,4-xylanase; endo-1,4-xylanase; endo- β -1,4- xylanase; endo-1,4- β -D-xylanase; 1,4- β -xylan xylanohydrolase; β -xylanase; β -1,4-xylan xylanohydrolase; endo-1,4- β -xylanase; β -D-xylanase
EC number:	3.2.1.8
Reaction:	Catalyses the endohydrolysis of $(1\rightarrow 4)$ - β -D-xylosidic linkages in xylans

The hydrolysis reaction scheme for endo-1,4-beta-xylanase is available under its record in the enzyme database BRENDA² (Chang et al. 2021).

2.2 Manufacturing process

2.2.1 Production of the enzyme

Novozymes' endo-1,4-beta-xylanase is produced by submerged fermentation of a *T. reesei* microorganism expressing a endo-1,4-beta-xylanase enzyme from *T. leycettanus*. The manufacturing process begins with fermentation followed by formulation and then quality control for the enzyme preparation. The fermentation steps include preparation of the inoculum, seed fermentation followed by the main fermentation. The recovery stage includes multiple steps to separate, concentrate and stabilise the enzyme. Finally, the enzyme is blended into an enzyme preparation.³

The applicant states that the enzyme is manufactured in accordance with current Good Manufacturing Practice for Food (cGMP) and the principles of Hazard Analysis and Critical Control Point (HACCP). Novozymes follow relevant EC regulations for manufacture and packaging of food enzymes and for control and inspection. The quality management system complies with ISO9001:2015 for the development, production and sale of industrial enzymes at their Denmark facility.

The applicant states that all raw materials used in the production of the enzyme are of food grade quality that comply with relevant specifications.

Details on the manufacturing process, raw materials and ingredients used in the production of the endo-1,4-beta-xylanase enzyme preparation were provided in the application as Confidential Commercial Information (CCI).

² <u>https://www.brenda-enzymes.org/enzyme.php?ecno=3.2.1.8</u>

³ Enzymes are generally sold as enzyme preparations, which consist of the enzyme(s) and other ingredients, to facilitate their storage, sale, standardisation, dilution or dissolution.

2.2.2 Allergen considerations

The applicant provided the Product Data Sheet for the enzyme preparation. This states that certain allergens are not present including cereals containing gluten, crustaceans, egg, fish, lupin, milk (including lactose), molluscs, nuts, peanuts, sesame, soy, sulphur dioxide/sulphites. The applicant also sent additional information to FSANZ as CCI, providing supporting information regarding the absence of the identified allergens in their enzyme preparation.

2.2.3 Specifications

There are international specifications for enzyme preparations used in the production of food. These have been established by the Joint FAO/WHO Expert Committee on Food Additives (JECFA) in its Compendium of Food Additive Specifications (FAO/WHO 2006) and in the Food chemicals codex (FCC 2008). These specifications are included in earlier publications of the primary sources listed in section S3—2 of Schedule 3 of the Code and enzymes used as a processing aid must meet either of these specifications. The applicant states that the final enzyme preparation is consistent with the requirements in both specifications. Schedule 3 of the Code also includes specifications for arsenic and heavy metals (section S3—4) if they are not already detailed within specifications in sections S3—2 or S3—3.

The applicant provided the results of analysis of three representative batches of the xylanase preparation. Table 2 provides a comparison of this analysis with international specifications established by JECFA and Food chemicals codex, as well as those in the Code (as applicable). Based on these results, the enzyme preparation met all relevant specifications for arsenic, metals and the microbiological criteria.

Analysis	Analysis provided by manufacturer	FAO/WHO JECFA (2006)	Specifications Food Chemicals Codex (FCC, 2020)	Australia New Zealand Food Standards Code (section S3—4)
Lead (mg/kg)	<0.5	≤5	≤5	≤2
Arsenic (mg/kg)	<0.3	-	-	≤1
Cadmium (mg/kg)	<0.05	-	-	≤1
Mercury (mg/kg)	<0.05	-	-	≤1
Total Coliforms (CFU/g)	<4	≤30	≤30	-
Enteropathogenic <i>E.</i> <i>coli</i> (in 25 g)	Not detected	Absent	-	-
<i>Salmonella</i> spp. (in 25 g)	Not detected	Absent	Negative	-
Antimicrobial activity	Not detected	Absent	-	-

Table 2Comparison of Novozymes endo-1,4-beta-xylanase enzyme preparationcompared to FAO/WHO JECFA, Food chemicals codex and Code specifications for enzymes

While the manufacturing process ensures the production microorganism, *T. reesei* is absent from the final enzyme preparation, the food enzyme is a biological isolate of variable composition, containing the enzyme protein, as well as organic and inorganic material derived from the microorganism and fermentation process. Refer to section 2.3 for the total organic solids (TOS) value.

2.3 Technological purpose of the enzyme

Endo-1,4-beta-xylanase is intended for use in brewing, fats and oils processing, grain processing and potable alcohol production.

The enzyme performs its technological function by hydrolysing xylosidic linkages in xylans, including arabinoxylan present in grains for further processing to produce a range of products including gluten, starch, potable alcohol, beer, fats and oils. Hemicellulose is a complex mixture of polysaccharides of which xylans are the main component (Saha 2000). Given the complexity of its structure and the types of glycosidic linkages, complete hydrolysis of xylan requires what is termed 'accessory enzymes' - β -D-xylosidases, α -L-arabinofuranosidase, α -glucoronidase and esterases (Saha 2000; Yeoman 2010).

Information provided by Novozymes on the physical and chemical properties of their enzyme preparation is summarised in Table 3.

Physical and c	hemical properties of commercial enzyme preparation
Enzyme activity	1500 *FXU(S)/g
Appearance	Amber coloured liquid
Storage conditions	0–25°C
Density	1.24 g/mL
Viscosity	1-25 cps

 Table 3
 Physical and chemical properties of the endo-1,4-beta-xylanase preparation

*FXU(S)/g is Fungal Xylanase Units per gram

The enzyme preparation is available as a liquid concentrate standardised in fungal xylanase units to an activity of 1500 FXU(S)/g. Endo-1,4-beta-xylanases hydrolyse wheat arabinoxylan to release a reducing carbohydrate. The reaction is stopped by an alkaline reagent that complexes with the reducing sugar producing colour detected at 405nm. The increase in absorbance is proportional to the enzyme activity.

The highest dosage level used for solid and liquid foods is 600 FXU(S)/kg grains or 16mg TOS/kg grains.

The Codex guideline, *Guidelines on Substances used as Processing Aids* (CAC/GL 75-2010) sets out general principles for the safe use of substances used as processing aids. The Guideline states that substances used as processing aids shall be used under conditions of good manufacturing practice (GMP). Therefore, use of commercial enzyme preparations should follow GMP, where use is at a level that is not higher than that necessary to achieve the desired enzymatic reaction. The applicant has mentioned that use of the enzyme is at GMP levels.

Endo-1,4-beta-xylanase from *T. reesei* is already permitted to be used as a processing aid by the Code, but not from *T. reesei* containing for gene for endo-1,4-beta-xylanase from *T. leycettanus* as requested by the applicant.

2.4 Technological justification

The enzyme performs its technological function by hydrolysing xylosidic linkages in xylans, including arabinoxylan present in grains for further processing. The enzyme is denatured and inactivated during brewing, fats and oils processing, grain processing and potable alcohol production. It therefore functions as a processing aid for the purposes of the Code.

The applicant has provided information on claimed benefits associated with the use of endo-1,4-beta-xylanase in brewing, fats and oils and grain processing, and potable alcohol

production. These are summarised in Table 4.

Brewing	Fats and oils	Grains	Potable alcohol
Faster and known lautering or mash filtration	Higher oil extraction from same amount of raw material	Higher gluten and starch yield from efficient and targeted degradation of arabinoxylans	Higher solid conc. During mashing – more energy efficient
Increased options for raw material selection	Reduced viscosity during pressing and separation	Efficient water removal leading to energy savings	Improved heat exchange
Improved yield from efficient processing, less raw material use	Reduced oil loss	Efficient operations	Improved centrifugal separation
Faster beer filtration	Reduced water use	Overall reduced grain cost	Improved mass transfer during fermentation
Reduced use of filtration aids	Reduced waste production and handling		Increased fermentable sugars following beta glucan hydrolysis

Table 4 Benefits associated with use of endo-1,4-beta-xylanase in food produc

2.5 Food technology conclusion

FSANZ concludes that the proposed use of endo-1,4-beta-xylanase in brewing, fats and oils and grain processing, and potable alcohol production is consistent with its technological purpose of endo-hydrolysis of 1,4-beta-xylosidic linkages in xylan. FSANZ concludes that the information included in the application and assessment support the proposed use. The use of the enzyme, in the form and requested amount at a level consistent with GMP is technologically justified and has been demonstrated to be effective in achieving its stated purpose.

Endo-1,4-beta-xylanase performs its technological purpose during the production and processing of food and is not performing a technological purpose in the final food. It is therefore appropriately categorised as a processing aid as defined in the Code.

There are relevant identity and purity specifications for the enzyme in the Code and the applicant provided evidence that the enzyme meets these specifications.

3 Safety assessment

The objective of this safety assessment are to evaluate any potential public health and safety concerns that may arise from the use of this enzyme, produced by this microorganism, as a processing aid.

Some information relevant to this section is CCI, so full details cannot be provided in this public report.

3.1 History of use

3.1.1 Host organism

Trichoderma reesei

T. reesei is a common, hypercellulolytic, soil fungus that was initially isolated from deteriorating canvas made from cellulosic material. The original isolate QM6a is the type strain for *T. reesei* (Olempska-Beer et al. 2006) and has been registered with the American Type Culture Collection under ATCC13631. Strain QM6a is the wild type of practically all *T. reesei* industrial production strains (Nevalainenet et al. 1994). Due to the secretion of a range of cellulolytic enzymes, this fungus has been used since the 1980s for the industrial production of enzymes for a range of industries including food (Nevalainen and Peterson 2014; Paloheimo et al. 2016).

Several review papers support the safety of *T. reesei* with no production of known mycotoxins or antibiotics under conditions used for enzyme production (Nevalainen et al. 1994; Kubicek et al. 2007; Peterson and Nevalainen 2012; Frisvad et al. 2018). It is listed as Risk Group 1 in the microorganism classification lists of the German Federal Institute for Occupational Safety and Health (BAuA 2016) and meets the requirements of a Biosafety Level 1 organism based on the Biosafety in Microbiological and Biomedical Laboratories guidelines⁴.

FSANZ has previously assessed the safety of *T. reesei* as the source organism for 15 processing aids in Schedule 18. The *T. reesei* production strain in this application was developed from the well-known type strain QM6a. The *T. reesei* strain RUT-C30 (ATCC 56765) was developed from QM6a. The RUT-C30 strain is well known and documented (Peterson and Nevalainen 2012). The recipient strain used in the construction of the *T. reesei* production strain was derived from the RUT-C30 parental strain by a series of classical mutagenesis steps and spontaneous mutations. Data confirming the identity of the recipient strain was provided in the application.

The production organism is absent from the final enzyme preparation, the manufacturing process involves appropriate controls to prevent microbial contamination, and the microbial quality of the final enzyme preparation meets the specifications required by JECFA⁵. The production strain stability during fermentation was analysed using phenotypic parameters with no instability observed.

3.1.2 Gene donor organisms

Talaromyces leycettanus

The donor organism of the endo-1,4-beta-xylanase gene is a *T. leycettanus* strain and is a biosafety level 1 organism with no safety concerns.

3.2 Characterisation of the genetic modification(s)

3.2.1 Description of the DNA to be introduced and method of transformation

An expression cassette containing the endo-1,4-beta-xylanase gene was introduced into the genome of the host strain, *T. reesei*. The wild type endo-1,4-beta-xylanase gene is derived from *T. leycettanus* genomic DNA and placed under the control of a *T. reesei* promoter and

⁴ <u>https://www.cdc.gov/biosafety/publications/bmbl5/index.htm</u>

⁵ https://www.fao.org/food/food-safety-quality/scientific-advice/jecfa/jecfa-additives/enzymes/en/

terminator. Data provided by Novozymes and analysed by FSANZ confirmed the identity of the endo-1,4-beta-xylanase enzyme.

A hybrid *Saccharomyces cerevisiae/Escherichia coli* vector containing the endo-1,4-betaxylanase expression cassette was used to transform the host strain. The expression cassette was integrated at specific integration sites in the host's genome and the final production strain was selected based on rapid growth and high endo-1,4-beta-xylanase activity.

3.2.2 Characterisation of inserted DNA

Southern blot data provided by Novozymes and analysed by FSANZ confirmed the presence of the inserted DNA in the production strain. The applicant also provided the results of genome sequencing which confirmed the absence of antibiotic resistance genes in the production strain.

3.2.3 Genetic stability of the inserted gene

The assessment confirmed the inserted gene is integrated into the genome of the production strain and does not have the ability to replicate autonomously. The inserted gene is therefore considered to be genetically stable.

To provide further evidence of the stability of the introduced gene, the applicant provided phenotypic data from large-scale fermentation of the production strain. These data confirmed that the gene encoding the endo-1,4-beta-xylanase enzyme is expressed over multiple generations and is stable.

3.3 Safety of endo-1,4-beta-xylanase

3.3.1 History of safe use

Multiple xylanase enzymes are currently permitted as processing aids in Schedule 18 of the Code. However, *T. leycettanus* endo-1,4-beta-xylanase is not permitted and does not have a history of safe use in Australia or New Zealand. The applicant stated that the enzyme is approved for use in Denmark (Jan 2019), Mexico and Brazil.

There are no known reports of adverse effects arising from the consumption of endo-1,4beta-xylanase, nor *T. leycettanus* endo-1,4-beta-xylanase, when used as a processing aid.

3.3.2 Bioinformatics assessment for enzyme toxicity

A Clustal alignment (<u>ClustalW 2.0.10</u>⁶) was performed by the applicant comparing the endo-1,4-beta-xylanase protein sequence against all sequences in the <u>UniProt</u>⁷ database (release: 2021-02-15) that contained the search word 'toxin' and not 'fragment'.

There were no toxin sequences identified with a sequence similarity to the enzyme above 20% identity, indicating that the endo-1,4-beta-xylanase enzyme has no substantial homology with known protein toxins.

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Clustal W2 available from: <u>http://www.clustal.org/clustal2</u>

⁷ UniProt database: <u>https://www.uniprot.org</u>

3.3.3 Evaluation of toxicity studies

The *T. leycettanus* endo-1,4-beta-xylanase test item used in the supplied toxicity studies was produced using GM *T. reesei*, and was equivalent to the enzyme product for commercialisation.

Animal studies

<u>90-day repeated dose oral toxicity study in rats (Envigo CRS Limited, 2016). Regulatory</u> <u>Status: GLP; conducted according to OECD Test Guideline (TG) 408.</u>

The endo-1,4-beta-xylanase test item was administered to Wistar Han rats (10/sex/group) at doses of 0, 105, 347, 1051 mg total organic solids (TOS)/kg body weight (bw)/day by oral gavage for 13 weeks. The vehicle control was water.

Animals were observed twice daily, with detailed observation performed twice weekly. Body weight, food consumption and detailed physical examinations for signs of toxicity were recorded weekly. Ophthalmological examination was conducted on all test animals prior to treatment and on the control and high-dose animals at study termination. Functional performance and sensory reactivity tests were performed in week 12. Gross pathology, haematology, clinical chemistry and measurement of organ weights was conducted on all animals at termination. Histopathological examination was conducted on the control and high-dose test groups only.

There was no treatment-related mortality during the study period, nor treatment-related effects on feed consumption, body weights, haematology, clinical chemistry, ophthalmology, or functional observations (functional performance or sensory reactivity). No treatment-related macroscopic abnormalities or histopathological findings were observed in any of the test animals at necropsy.

The No Observed Adverse Effect Level (NOAEL) was identified as 1051 mg TOS/kg bw/day, which was the highest dose tested.

Genotoxicity

Bacterial reverse mutation test (Covance Laboratories Ltd, 2016). Regulatory Status: GLP; conducted according to OECD TG 471.

The potential mutagenicity of endo-1,4-beta-xylanase was evaluated in *Salmonella enterica* ser. Typhimurium strains TA98, TA100, TA1535, TA1537 and *Escherichia coli* strain WP2 *uvrA* pKM101, with and without metabolic activation using rat liver homogenate (S9). Bacterial cultures were treated for 1 hour with the test item between $16 - 5000 \mu g$ TOS/mL, before treatment was removed and bacterial cultures plated using a modified treat-and-plate method. The experiment was conducted twice independently.

Positive controls in the absence metabolic activation were 2-nitrofluorene (TA98), 4nitroquinoline 1-oxide (TA100), *N*-methyl-*N*-nitro-*N*-nitrosoguanidine (TA1535, WP2 *uvrA* pKM101) and ICR-171 (TA1537). The positive control in the presence of metabolic activation was 2-aminoanthracene (all strains). Purified water was used as the vehicle control.

No concentration-related increases in revertant colonies were observed in cultures treated with the test item, relative to vehicle controls, with or without metabolic activation. All positive control treatments showed the anticipated increases in mutagenic activity demonstrating the validity of the assay.

It was concluded that endo-1,4-beta-xylanase test item was not mutagenic under the conditions of this test.

In vitro mammalian micronucleus test in human lymphocytes (Covance Laboratories Ltd, 2016). Regulatory status: GLP; conducted according to OECD TG 487.

The potential of endo-1,4-beta-xylanase to induce micronuclei formation in mammalian cells was tested using human lymphocytes isolated from peripheral blood, collected and pooled from two healthy male volunteers. Treatment with the endo-1,4-beta-xylanase test item was either a 3-hour pulse exposure with or without S9, followed by a 21-hour recovery; or 24 hours of exposure with without S9, followed by 24-hour recovery. Positive control assays were conducted in parallel using mitomycin C as the clastogen positive control in the pulse treatment without S9, cyclophosphamide as the clastogen positive control in the pulse treatment with S9, and vinblastine as the aneugen positive control in the 24-hour exposure without S9. Results were an average of two experimental replicates. Sterile water was used as the vehicle control.

As a result of dose-selection experiments, the dose range for all treatments were adjusted to $500 - 5000 \ \mu g \ TOS/mL.$

There were no treatment-related increases in the incidence of micronucleated human lymphocytes following exposure to the test item, relative to the vehicle controls, under the conditions tested. The positive controls demonstrated a statistically significant increase in micronuclei formation, validating the sensitivity of the experimental methodology. It was concluded that the test endo-1,4-beta-xylanase was not clastogenic or aneugenic in human lymphocytes, under the conditions of this study.

3.3.4 Potential for allergenicity

A FASTA search was performed using the amino acid sequence of endo-1,4-beta-xylanase using the <u>AllergenOnline</u>⁸ database (queried April 2021) using four sequence alignments: the full-length protein (more than 35% identity), an 80 mer sliding window (more than 35% identity), a scaled 80 mer sliding window (more than 35% identity) and an 8 mer sliding window (100% identity). No allergen sequences were identified using these search parameters.

Xylanases are known respiratory allergens arising from occupational sensitisation when used in industrial settings, such as commercial baking or animal feed settings (Merget et al. 2001; Vanhanen et al. 2001; Elms et al. 2003). However, food enzyme processing aids that are respiratory allergens are not usually food allergens (Poulsen 2004, Bindslev-Jensen et al. 2006) and FSANZ did not locate any case reports in the scientific literature of oral sensitisation to xylanase enzymes.

3.3.5 Assessments by other regulatory agencies

The *T. leycettanus* endo-1,4-beta-xylanase produced using GM *T. reesei* is approved for use in Denmark, Mexico and Brazil. The written assessments undertaken by these countries were not provided to FSANZ.

3.4 Dietary exposure assessment

The objective of the dietary exposure assessment was to review the budget method calculation presented by the applicant as a 'worse-case scenario' approach to estimating likely levels of dietary exposure, assuming that all of the TOS from the endo-1,4-beta-xylanase enzyme preparation remained in the food.

⁸ AllergenOnline: <u>http://www.allergenonline.org/</u>

The budget method is a valid screening tool for estimating the theoretical maximum daily intake (TMDI) of a food additive (Douglass et al. 1997). The calculation is based on physiological food and liquid requirements, the food additive concentration in foods and beverages, and the proportion of foods and beverages that may contain the food additive. The TMDI can then be compared to an ADI or a NOAEL to estimate a margin of exposure for risk characterisation purposes. Whilst the budget method was originally developed for use in assessing food additives, it is also appropriate to use for estimating the TMDI for processing aids (FAO/WHO 2020). The method is used by international regulatory bodies and the FAO/WHO Joint Expert Committee on Food Additives (JECFA) (FAO/WHO 2021) for dietary exposure assessments for processing aids.

In their budget method calculation, the applicant made the following assumptions:

- the maximum physiological requirement for solid food (including milk) is 25 g/kg body weight/day
- 50% of solid food is processed
- on average, all processed food contains 25% grains (or grain-derived) dry matter
- the maximum physiological requirement for liquid is 100 mL/kg body weight/day (the standard level used in a budget method calculation for non-milk beverages)
- 25% of non-milk beverages are processed
- all processed beverages contain 12% grain hydrolysates
- densities of the non-milk beverages are ~ 1
- all solid foods and non-milk beverages contain the highest use level of 16 mg TOS/kg raw material (grains)
- all of the TOS from the enzyme preparation remains in the final food
- Based on these assumptions, the applicant calculated the TMDI of the TOS from the enzyme preparation to be 0.1 mg TOS/kg body weight/day.

As assumptions made by the applicant differ from those that FSANZ would have made in applying the budget method, FSANZ independently calculated the TMDI using the following assumptions that are conservative and reflective of a first tier in estimating dietary exposure:

- The maximum physiological requirement for solid food (including milk) is 50 g/kg body weight/day (the standard level used in a budget method calculation where there is potential for the enzyme preparation to be in baby foods or general purpose foods that would be consumed by infants).
- FSANZ would generally assume 12.5% of solid foods contain the enzyme based on commonly used default proportions noted in the FAO/WHO Environmental Health Criteria (EHC) 240 Chapter 6 on dietary exposure assessment (FAO/WHO 2009).

However, the applicant has assumed a higher proportion of 50% based on the nature and extent of use of the enzyme and therefore FSANZ has also used this proportion for solid foods as a worst-case scenario.

All other inputs and assumptions used by FSANZ remained as per those used by the applicant. The TMDI of the TOS from the enzyme preparation based on FSANZ's calculations for solid food is and non-milk beverages is 0.15 mg TOS/kg body weight/day.

Both the FSANZ and applicant's estimates of the TMDI will be overestimates of the dietary exposure given the conservatisms in the budget method. This includes that it was assumed that all of the TOS from the enzyme preparation remains in the final foods and beverages whereas the applicant has stated that it is likely to either be reduced or removed during processing or would be present in insignificant quantities. In addition, the enzyme would be inactivated and perform no function in the final food to which the ingredient is added.

4 Discussion

No public health and safety concerns were identified in the assessment of endo-1,4-betaxylanase produced by this GM *T. reesei* under the proposed use conditions. *T. reesei* has a long history of safe use as a production microorganism of enzyme processing aids, including several that are already permitted in the Code. The production organism is neither pathogenic nor toxigenic. Analysis of the modified production strain confirmed the presence and stability of the inserted DNA. Bioinformatics analysis indicated that the produced endo-1,4-beta-xylanase does not demonstrate substantial homology with any known toxins or food allergens.

Toxicity testing of the endo-1,4-beta-xylanase enzyme showed no evidence of genotoxicity *in vitro* and the NOAEL in a 90-day oral gavage study in rats was the highest dose tested, 1051 mg TOS/kg bw/day. The TMDI based on FSANZ's calculations is 0.15 mg TOS/kg body weight/day. Comparison of the NOAEL and the TMDI results in a Margin of Exposure (MOE) of approximately 7000.

5 Conclusion

Based on the reviewed data it is concluded that in the absence of any identifiable hazard an Acceptable Daily Intake (ADI) 'not specified' is appropriate.

6 References

Bindslev-Jensen C, Skov PS, Roggen EL, Hvass P, Brinch DS (2006) Investigation on possible allergenicity of 19 different commercial enzymes used in the food industry. Food Chem Toxicol. 44(11):1909-15

Bundesanstalt für Arbeitsschutz und Arbeitsmedizin (BAuA) (2016) Technische Regel für Biologische Arbeitsstoffe 460. <u>https://www.baua.de/DE/Angebote/Rechtstexte-und-</u> <u>Technische-Regeln/Regelwerk/TRBA/TRBA-460.html</u>

Chang A, Jeske L, Ulbrich S, Hofmann J, Koblitz J, Schomburg I, Neumann-Schaal M, Jahn D, Schomburg D (2021) BRENDA, the ELIXIR core data resource in 2021: new developments and updates. Nucleic Acids Res. 49:D498–D508. DOI: 10.1093/nar/gkaa1025

Covance Laboratories Ltd (2016) Xylanase, batch PPQ40100: Bacterial Reverse Mutation Assay using a Treat and Plate Modification. *Study Number:* 8336499

Covance Laboratories Ltd (2016) Xylanase, batch PPQ40100: *In Vitro* Human Lymphocyte Micronucleus Assay. *Study Number:* 8336501

Douglass JS, Barraj LM, Tennant DR, Long WR and Chaisson CF (1997) 'Evaluation of the Budget Method for screening food additive intakes, Food Additives and Contaminants' 14:791-802.

Elms J, Fishwick D, Walker J, Rawbone R, Jeffrey P, Griffin P, Gibson M, Curran AD. (2003) Prevalence of sensitisation to cellulase and xylanase in bakery workers. Occup Environ Med. 60(10):802-4.

Envigo CRS Ltd (2016) Arabinofuranosidase, Batch PPH40331: Xylanase, Batch PPQ40100: Toxicity Study by Oral Gavage Administration to Han Wistar Rats for 13 Weeks. *Study Number: DK60TW* FAO/WHO (2009) 'Environmental Health Criteria 240. Principles and Methods for the Risk Assessment of Chemicals in Food' Chapter 6 – Dietary exposure assessment of chemicals in food, WHO, Geneva.

FAO/WHO (2020) Environmental Health Criteria 240. Principles and Methods for the Risk Assessment of Chemicals in Food. Chapter 6: Dietary exposure assessment of chemicals in food. Second Edition 2020. WHO, Geneva. <u>https://www.who.int/docs/default-source/food-safety/publications/chapter6-dietary-exposure.pdf?sfvrsn=26d37b15_6</u>

FAO/WHO (2021) Evaluation of certain food additives: eighty-ninth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series, No. 1027.

FAO/WHO (Food and Agriculture Organization of the United Nations/World Health Organization) (2006) General specifications and considerations for enzyme preparations used in food processing in Compendium of food additive specifications. 67th meeting. FAO

JECFA Monographs, 3, 63–67. Available online: <u>Compendium of Food Additive</u> <u>Specifications. Joint FAO/WHO Expert Committee on Food Additives (JECFA), 67th meeting</u> <u>2006. FAO JECFA Monographs 3</u>FCC (2020) Enzyme preparations. In: Food Chemicals Codex, 12th edition. Rockville (MD): United States Pharmacopeial Convention.

Frisvad JC, Møller LL, Larsen TO, Kumar R, Arnau J. (2018) 'Safety of the fungal workhorses of industrial biotechnology: update on the mycotoxin and secondary metabolite potential of Aspergillus niger, Aspergillus oryzae, and Trichoderma reesei', *Applied Microbiology and Biotechnology*, 102(22):9481-9515.

IUBMB (2022) EC 3.2.1.8 (gmul.ac.uk) (website last updated July 2022)

Kubicek CP, Komoń-Zelazowska M, Sándor E, Druzhinina IS (2007) 'Facts and Challenges in the Understanding of Biosynthesis of Peptaibols by Trichoderma', *Chemistry and Biodiversity*, 4(6):1068-1082.

McDonald AG, Boyce S, Tipton KF (2009) ExplorEnz: the primary source of the IUBMB enzyme list. Nucleic Acids Res 37:D593–D597 (2009). DOI: 10.1093/nar/gkn582

Merget R, Sander I, Raulf-Heimsoth M, Baur X. (2001) Baker's asthma due to xylanase and cellulase without sensitization to alpha-amylase and only weak sensitization to flour. Int Arch Allergy Immunol. 124(4):502-5.

Nevalainen H and Peterson R (2014) 'Making recombinant proteins in filamentous fungi - are we expecting too much?', *Frontiers of Microbiology*, 5:e75.

Nevalinen H, Suominen P, Taimisto K (1994) 'Minireview. On the Safety of Trichoderma Reesei', *Journal of Biotechnology*, 37:193-200.OECD (2014). 'Genetic Toxicology: OECD Guideline for the testing of chemicals. Guideline 487: *In vitro* mammalian cell micronucleus test

Olempska-Beer ZS, Merker RI, Ditto MD and DiNovi MJ (2006) 'Food-processing enzymes from recombinant microorganisms – A review' *Regulatory Toxicology and Pharmacology* 45:144-158.

Paloheimo M, Haarmann T, Mäkinen S and Vehmaanperä J (2016) 'Production of industrial enzymes in Trichoderma reesei.' In: Schmoll M, Dattenböck C (Eds) Gene expression systems in Fungi: Advancements and Applications, Springer International Publishing, Switzerland, p.23-57.

Peterson R and Nevalainen H (2012) 'Trichoderma reesei RUT-C30-thirty years of strain improvement', *Microbiology*, 158: 58-68.

Poulsen LK (2004) Allergy assessment of foods or ingredients derived from biotechnology, gene-modified organisms, or novel foods. Mol Nutr Food Res. 48(6):413-23

Saha BC (2000). Research Review paper: α-L-Arabinofuranosidases: biochemistry, molecular biology and application in biotechnology. Biotechnology Advances 18(5), 402-423

Seidl V, Gamauf C, Druzhinina IS, Selboth B, Hartl L, Kubicek CP (2008) 'The Hyocrea jecorina (Trichoderma reesei) hypercellulolytic mutant RUT C30 lacks a 85 kb (29 geneencoding) region of the wild-type genome', *BMC Genomics* 9:327-342.

Vanhanen M, Tuomi T, Tiikkainen U, Tupasela O, Tuomainen A, Luukkonen R, Nordman H (2001) Sensitisation to enzymes in the animal feed industry. Occup Environ Med. 58(2):119-23.

Yeoman CJ, Han Y, Dodd D, Schroder CM, Mackie RI and Cann IKO (2010) Chapter 1 Thermostable Enzymes as Biocatalysts in the Biofuel Industry. In: Advances in Microbiology, Volume 20, p. 19.